



Journal of Integral Sciences (JIS)

(An International Open Access Journal)

Available at www.jisciences.com

ISSN: 2581-5679

A review on bioanalytical method development and validation of anticancer drugs by using LC/MS/MS and its applications on routine analysis

Anil Kumar Tallam¹, Alapati Sahithi², Mohana Vamsi Nuli³

¹Department of pharmacy, Shri Venkateshwara University, Rajabpur, NH-24, Venkateshwara Nagar, Gajraula, Uttar Pradesh 244236

²Assistant Professor, Department of Pharmaceutical Analysis, Nalla Narasimha Reddy Education Society's Group of Institutions, Narapally, Ghatkesar Mandal, Korremula Rd, Hyderabad, Telangana 500088

³Associate Professor, Raghavendra Institute of Pharmaceutical Education and Research K.R. Palli Cross, Dist Anantapuramu, Chiyvedu, Andhra Pradesh 515721

Received: 22 Dec 2022 Revised: 04 Jan 2023 Accepted: 13 Feb 2023

Abstract

A protocol is used to detect and measure biomolecules and metabolites in human and animal tissues using bimolecular methods. The biosanality method is effective at determining the number of drugs and metabolites in a biological system. New methods, the validation of existing procedures, and the analysis of samples are one of the prominent tasks for bioanalysis. Above all, a compound can be measured using several methods and identified by different methods of analysis. Drugs may be tested by several extraction techniques, including liquid extraction, solid-phase extraction, and protein precipitation in complex plasma and biological samples. To determine how the environment, matrix, or procedures impact the matrix estimation to the time of the analysis, all steps in the process must be investigated. The more detailed study of drug products can be performed with higher-pressure analytical techniques, such as high-extraction (HPLC), liquid chromatography coupled with double-mass spectrometry (LCMS/MS), and ultra-performance Liquid chromatography (UPLC). Both of them have flaws and strengths. At present, HPLC and GC usually perform biolysis. The parameters are linearity, repeatability, accuracy, selectivity, and continuity. We are proposing the development and validation of bioanalytical systems to assist in the quality assurance of drugs.

Keywords: Bioanalytical, samples extraction techniques, LC-MS/MS, UPLC.

This article is licensed under a Creative Commons Attribution-Non-commercial 4.0 International License.

Copyright © 2022 Author(s) retains the copyright of this article.



*Corresponding Author

Anil Kumar Tallam

DOI: <https://doi.org/10.37022/jis.v6i1.51>

Produced and Published by

[South Asian Academic Publications](#)

Introduction

The application of the bioanalytical approach is one of the drug developments. To quantitatively determine different analytes in biological matrices, bioanalytical method validation is essential. Good preparation of samples and hyphenated instruments is crucial in modern bioanalysis. The development of complete bioanalytical methods in pharmaceutical research companies is critical during the drug discovery and development phase [1]. Bioanalysis covers identifying and quantifying biological sample

analytes (blood, plasma, serum, saliva, urine, feces, skin, hair, organ tissue). In Bioanalysis, small molecules like drugs and metabolites are met, whereas large molecules like proteins and peptides are identified. Bioanalysis is an essential science in many fields of study, such as developing new medicines, forensic testing, doping controls, and the identification of biomarkers for many diseases. Due to the sophistication of the sample matrix, Bioanalysis is challenging [2-11]. There is a well-known need for intense sample preparation before being used in analytical instruments to use complex matrices such as blood, plasma, and urine. Modern bioanalysis requires high-performance sample preparation and hyphenated analytical methods. For a long time in bioanalysis, medicine has been used with liquid (LC) paired to tandem mass spectrometry (MS/MS).

Gemcitabine hydrochloride

Gemcitabine hydrochloride is a β -difluoro-nucleosides, purine antimetabolite (4-amino-1-[(2R, 4R, 5R)-3, 3-difluoro-4-hydroxy-5-(hydroxymethyl) oxolan-2-yl] pyrimidin-2-one [12]. A mixture of the diphosphate and the triphosphate nucleosides leading to inhibition in DNA synthesis Gemcitabine [13]. It was first approved by the FDA in 1996 and has been shown for breast, ovary, non-small cell lung, and pancreatic cancer [14]. Gemcitabine HCL is water-soluble, methanol-soluble, and ethanol and polar organic solvents are virtually insoluble [15]. It is white and odorless crystalline and powder with a 168.64°C melting point. Gemcitabine's molecular formula is $C_9H_{11}F_2N_3O_4$, and 263.2 g/mol is molecular [16]. It inhibits the growth of tumors by two mechanisms: first through the replacement of one nucleic acid building block directly during DNA replication, leading to tumor cell apoptosis, and second, by irreversibly inactivating the ribonucleotide reductase enzyme, which prevents the development of deoxyribonucleotide and causes cell apoptosis [17]. Gemcitabine is also used in pancreatic adenocarcinoma. Patient supervision shows only a small improvement and its efficacy can be restricted by the poor drug administered, particularly for pancreatic adenocarcinoma tumors which are usually hypo-vascular and extensive desmoplastic stroma [18]. Gemcitabine has a half-life of 17 minutes [19].

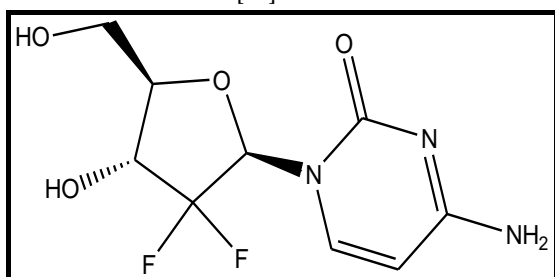


Fig-1 structure of Gemcitabine hydrochloride

Dasatinib

Dasatinib is a multi-tyrosine kinase inhibitor. Dasatinib has blocked the expression of BCR-AB1 by developing chronic leukemia myeloid and acute lymphoblastic leukemia lines [20]. The IUPAC of Dasatinib N-(2-chloro-6-methyl phenyl)-2-[[6-[4-(2-hydroxyethyl) piperazin-1-yl]-2-methylpyrimidin-4-yl] amino]-1, 3-thiazole-5-carboxamide with an empirical formula of $C_{22}H_{26}ClN_7O_2S$ with a molecular weight of 488.01 g.mol⁻¹. Dasatinib is slightly soluble and very poorly soluble in ethanol, methanol, DMSO, acetone, acetonitrile [21]. Dasatinib has a melting point of 280°-286°C and white or off-white powder (22). The recommended starting dose of Sprycel is 100 mg given orally once daily for the chronic

phase [23]. Dasatinib has various strengths such as 20 mg, 50 mg, 70 mg, 80 mg, 100 mg, and 140 mg available on the market [24]. Dasatinib inhibits the kinase BCR-ABL, SRC (SRC, LCK, YES, FYN), C- KITA, EPHA2, and pdgfr β at Nano molar concentrations. Based on modeling studies, Dasatinib is expected to be associated with several ABL kinase conformations [25].

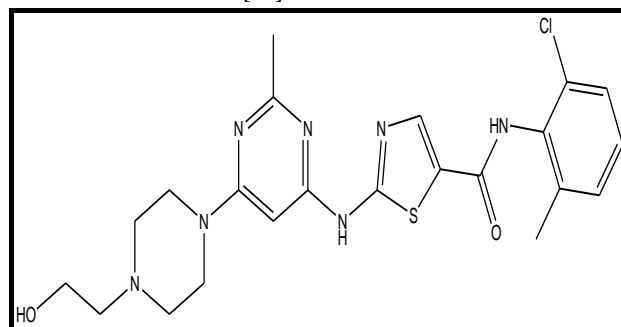


Fig-2 Structure of DasatinibRuxolitinib

Ruxolitinib is an active, selective inhibitor indicated for the treatment of moderate-and high-risk myelofibrosis, including primary icelophibrosis, post-polycythemia, and post-essential thrombocythemia myelofibrosis. The food and drug administration (FDA) approved Ruxolitinib in 2011. [26]. Ruxolitinib with a (3R)-3-cyclopentyl-3-[4-(7H-pyrrolo [2,3-d] pyrimidin-4-yl) pyrazol-1-yl] propane nitrile molecular formulation, known chemically as (r)-3-3-(4-(7h-pyrrolo [2,3-d] [27]. When JAK2 mutations were detected in myelofibrosis, the emphasis was moved towards selective JAK inhibitors to control diseases. Ruxolitinib is a potent, first-class, and selective FDA-approved inhibitor for myelofibrosis therapy. A phase 1/2 clinical study is currently in progress to evaluate the impact of the Ruxolitinib/nilotinib combination in CML [28]. In this case, the cost of the National Health Service, the Italian medicine agency, also approved off-label use of diseases other than covid-19. In particular, for patients with diagnosed covid 19 and severe/severe lung disease, Ruxolitinib is part of the compassionate application of AIFA-approved medicines [29]. It is soluble with the inert gas in organic solvents such as ethanol, DMSO, and DMF [30].

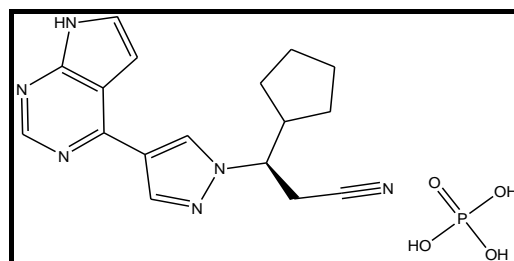


Fig-3 Structure of Ruxolitinib

Sample preparation

In terms of time and the complexity of extracting the desired analyte from the matrix, the analysis's sample preparation phase is often the most important and complicated part. Moreover, there are unique obstacles to each matrix. Urine has high levels of salt; for example, plasma has plenty of phospholipids. Whole blood comprises red, lysed blood cells, and so on. Each analyte and matrix often have different properties, which dictate the type of extraction method to use [31].

The compound detection

A high-performance liquid chromatography detector is a mass spectrometer combined with tandem-mass [HPLC-MS/MS] technology that utilizes both electrospray ionization [ESI] and Air Pressure Chemicals Ionization [APCI] technology [32,33]. Recent technological advancement has made material for the packaging of 1.7 μm particles available. UPLC, combined with a high-pressure pump and MS high-speed acquisition, offers extremely high efficiency, high-resolution capacity for optimum chromatography performance. [34].

Sample preparation goals [35]

Reduce the impact of matrix

Remove sample for variability sample

Reduce the variability of assays

Enhanced sensitivity

Samples should be cleaner

Understanding the challenges in method development [36]

The matrix

The number of specimens

Fast procedure

The quantification number of analyses

Analyte pharmaceutical profile

Ratio blood-to-plasma

A drug's blood to plasma ratio is its proportion to the corresponding plasma volume, i.e., CB/CP, in whole blood [containing red blood cells, RBCs, and plasma] [37].

Economic Aspects of Larger Molecule Drug Development

It takes approximately 10 years and takes about \$1 billion to find and produce a new drug. The average annual approval of large molecules is nine and twenty-three small molecules per year. Drug failures at various stages of production and patent expiry periods, primarily for existing organic products, contributed to a pharmaceutical R&D recession, as biologics are expected to reach a market value of \$54 billion off-patent in the next five years. Pharmaceutical companies are working

hard to reduce their internal capacities and fixed costs, and bioanalytical research the need to make bioanalytical sound tests help as a key tool for drug discovery and production must be well known, adding to the list of significant pharmaceutical expenditure. [38].

Bioanalysis and the different stages of drug development are generally recognized as central to the pharmacokinetic/Pharmacodynamic characterization of the novel chemical entity since it was discovered. Thus it led to its market authorization. There are a few general ideas in this field that are the foundation of an overarching structure for approaching bioanalysis from the outset and through different phases of drug production [39].

Extraction Methods

Liquid-Liquid Extraction

It is dependent on the selective extraction of the material from the liquid using the organic immiscible solvent of the analyte found in it. Solvents used include various hydrocarbons, ethylene dichloride, and diethyl acetate. Liquid-liquid extraction must provide differential solubility and then spread two immiscible fluids. The two phases had to be inseparable. In general, there will be two stages, one aqueous and the other containing living organisms. If required, the extraction step can be removed from the matrix.

Solid-Phase Extraction

It is based on the adsorption phenomenon with a particulate material [adsorbent] to which a specific substance is partitioned under a given set of conditions or selective adsorption of an analyte by a solid adsorbent. The Solid-phase Extraction function depends on the choice of adsorption. A target analyte can be extracted and/eluted using a preferred solvent [40].

Reverse-Phase SPE

The nonpolar [versatile or immobile] phase is brought into contact with a polar [mobile] liquid or solid phase. In the sample, the analyte is mid-to-low polar. They are mainly found in the metal industry but can also be used as lubricants. The use of semi-critical agents such as essential oils, barbiturates, vitamins, and phenols [C₁₈-octadecyl bonded for nonpolar to semi-polar extraction of antibiotics, benzodiazepines, nutrients, and drugs for use in polar beverages] [41].

Normal phase SPE

A mid-to-nonpolar polar analysis is used in normal-phase SPE [e.g., acetone, chlorinated solvents, and hexane]. Under normal circumstances, functionalized

silica [LCNH₂, NH₂, OR CN] is often used [LC- SI, SLC-alumina].

Ion exchange SPE

The exchange of positive/negative ions SPE is beneficial to solutions. Silicones are available with LC- SAX or LC-NH₂ connections. The LC-SCX or LC-WCX silica bonded silica cartridges are used to differentiate between cationic and anionic compounds.

Advantages of SPE OVER LLE [42]

Increased recovery and precision

Simplification in terms of automation,

Complete analyte availability

Protein precipitation

In a biological matrix, for example, an analyte, such as glucose, the solubility would be dependent on the primary solvent, e.g., plasma. Methanol and ACN are solvents used. It completely denatures the proteins. The majority of the protein may be precipitated with urea.

To change the PH and organic solvents are added to the sample. Since it's hydrophobic, it leads to spoiling the protein. The most recent developments are in sampling and tuning.

Salting-out assisted LLE

The organic solution is removed from the mix when salt is added to water. It is commonly regarded as "the salt-induced separation mechanism." nonpolar organic solvents may be extracted by the salting process.

Magnesium sulfate, ammonium sulfate sodium, and calcium sulfate [both dissolved and precipitated] were used in the process for treatment. The salting-out can be done utilizing an assisted LLE process that is inexpensive and can be mechanized [44].

Protein precipitation plates

Protein precipitation is done with a device called Protein precipitation, which allows filtration without subsequent centrifugation and supernatant transfer procedures. Although this series of filters has a membrane depth, they can be fastened to the vacuum system. Many samplers can easily be automated using the filtering plate as a support for the entire plan.

An Acetonitrile solvent is used. Protein precipitation filters should be chosen depending on the vapor pressure, pores, pore size, media, and other possible parameters, including solubility or matrices must be taken into consideration

Solid-phase extraction technique

SPE is similar to liquid chromatography in that it is based on the principle of affinity. [45]. SPE is:- recently Molecularly imprinted polymer Solid-phase extraction

In this method, the combination of template and functional monomers is formed by in-situ by noncovalent interactions such as electrostatic forces, Hydrogen binding, or vanderwaals force molecularly-imprinted polymer SPE [46] polymer

Molecularly imprinted polymer SPE applications: -

Analysis of the environment sample

Analysis of food and samples

Analysis of veterinary sample

Dispersive solid-phase extract

The solvent can be removed from a sample by organic solvents such as magnesium sulfate separation, acetonitrile acetone, magnesium sulfate separation or in conjunction with other salts [such as sodium chloride], and cleaned up by a dispersive solid-phase extract. Traditionally, the dispersive solid-phase extract approach is also used in [44]:

Sample tests of pesticides

Analysis of the food sample

Analysis of the environmental sample

Analyzes of cosmetic additives

Recent Dispersive solid-phase applications

Removal of Trace Elements

Pig tissue agonists

B-lactam antibiotic quantitative examination for the kidney tissue

Disposable pipette extraction

It is a dispersible sorbent powder that is free-flowing and contains a standard pipette tip [1/5 ml]. As a regular tip, it's simple to introduce and remove solvent from this scattering absorbent. A dispersible sorbent is positioned loosely between two modified frits [one at the tip's lower end, from which the solvent can be removed, and the other at the tip's top to prevent contamination by pipette solvents]. [44].

Micro-extraction by packed sorbent

1 mg sorbent packed into a 100-250 ml syringe and attached as a cartridge between the barrel and needle in micro-extraction using packed sorbent in the form of a syringe. Pre-treatment of complex sample sites, including dilution and centrifugation, is included in the micro-extraction by packed sorbent protocol. Dilution with centrifugation is recommended in the ratios of 1:4 and 1:20 for plasma/serum and whole blood samples, respectively. After collecting the sample, a Micro-extraction by packed sorbent syringe can be used to draw it. The analysts are tethered to the sorbent as the sample passes through the Meps cartridge. If the sample is needed before concentration, it can be extracted and expelled multiple times from the same vial to increase

the procedure's sensitivity. Following that, the sorbent bed is cleaned. The final step is to use a solvent that is sufficiently soluble to elude analytes. The elution method requires between 20 and 50 milliliters of organic solvent. Elution can be done directly in the injector of the instrument [GC or LC]. Meps can be used in bio-matricidal applications on plasma, serum, urine, whole blood, hair, and saliva.

Solid-phase micro-extraction

Solid-phase micro-extraction, which utilizes a fused silica fiber with an adequate stationary point on the outside, is a modern sample preparation technique. Physical calibration of the syringe is included, as is stainless steel micro tubing in the needle. This micro tubing has an organically polymerized fused silica

Sorptive extraction with a stir bar

The stir bar is equipped with a magnetic glass jacket. The glass jacket contains a given thickness layer of polymer. The test matrix is separated into the extraction process by analytes, as this bar is applied to the prepared sample. The total fiber coated Polydimethylsiloxane volume for solid-phase mining is 0,5 ml [film thickness of 100 nm] but in Stir bar sorptive mining it is 50–250 times greater. There was a 100mm thick film. Higher extraction levels are expected to produce greater extraction efficiency in stir bar-sorptive extraction than in solid-phase micro extraction. The spray extraction bar has two forms of extraction, one of which is SPME compatible, namely direct dipping and extraction of headspace.

On-line solid-phase extraction

Two kinds of online solid-phase columns are available for sale with a restricted access medium [ram] and a turbulent flow chromatograph column. Rams are often used in complex, high molecular material matrices to analyze low molecular mass substances [for example, medicine, endogenous substances, and xenobiotics] [most frequently proteins]. Columns of Ram HPLC remove the sample smoothing and are appropriate as a pre-column to guide biological, serum, and plasma samples. The mechanisms for the exclusion of hydrophilic, ion exchange, and scale are characterized as ram columns. Ram columns have been included in the multi-component analyses of biology-like antidepressant medicines [44]

Separation Liquid chromatography is a technique for partitioning a sample mixture into a column and separating it into two phases. The stationary phase within the column is the solvent, while the mobile phase that moves through the column is the solvent. Other chromatographic methods such as chromatography columns are often used in this thin layer chromatography. Detection of drugs in biological liquids using UV visible spectrometry fluorescence spectrometric [47].

Table -4 Extraction Values Of Gemcitabine

DRUG	MAT RIX	LC-COLUMN	SAMPLE PREPARATION	LLOQ	REFERE NCE
Gemcitabine	Human plasma	Acquity uplc hss T3 column (100A, 2.1 × 150 mm, 1.8 μm)	Solid-phase extraction	50,000Pg/ml	48
	Tumor tissue	Hypercarb column(100x 2.15μm)Thermo Fisher scientific)	Liquid-liquid extraction	0.2 ng/ml	49
	Human plasma	Altima c18 column(2.1×100 Mm, 5μm)	Protein precipitation	0.9 ng/ml	50
	Driedblood sample	BDS Hypersil c18,(100 x 4.6 mm, 5μ)	Protein precipitation	50 ng/ml	51
	Human peripheral Blood mononuclear Cells	A Biobasic 5 μm, 50 x 2.1 Mm column	Protein precipitation	25 ng/ml	52

Table -5 Extraction Values of Dasatinib

DRUG	MATRIX	LC – COLUMN	SAMPLE PREPARATION	LLOQ	REFER NCE
Dasatinib	Human plasma	Luna column (50 mm × 2.0 Mm 3-micron particle size)	Solid-phase extraction	0.92ng/ml	53
	Rat plasma	Acquity UPLC C ₁₈ analytical column (Waters, Dublin, Ireland), with dimensions 100 × 1.0 mm, i.d., 1.7 µm particle size.	Solid-phase extraction Liquid-liquid extraction	1.0ng/ml	54
	Rat plasma	Reversed-phase C ₁₈ column(50 4.6 mm i.d., 3 mm) YMC-Pack ODS-AM	Protein precipitation and liquid-liquid extraction	5.41 ng m ⁻¹	55
	Rat plasma	Reversed-phase C ₁₈ column(50 4.6 mm i.d., 3 mm) YMC-Pack ODS-AM	Liquid-liquid extraction	1.0ng/ml	56
	Rat plasma	Reverse phase C ₁₈ column (50 mm × 3 mm i.d., 4.6 µ) (YMC-PACK, Japan)	Protein precipitation	10 ng ml ⁻¹ .	57
	Human Plasma	Reversed-phase C ₁₈ column(50 4.6 mm i.d., 3mm) YMC-Pack ODS-AM	Solid-phase extraction	1 ng/ml	58
			Liquid-liquid extraction	62.5 ng/ml	49
			Liquid-liquid extraction	1 ng/ml	60
			Solid-phase extraction	0.1 ng/ml	61

Table-6 Extraction Values of Ruxolitinib

DRUG	MATRIX	COLUMN	SAMPLE PREPARATION	LLOQ	REFERENCE
Ruxolitinib	Human plasma	Phenomenex, synergi polar RP, 50X 2 mm, 4Mm	Liquid-liquid extraction	4.89 ng/ml	62
	Rat plasma	Ymc pack ods am (150 mm × 4.6 mm, 5 m)	Protein precipitation	0.86 ng/ml	63

Bioanalytical method validation

Need for validation of the bioanalytical process[64-72]

Accurate, reliable findings need to be satisfactorily interpreted by applying well-characterized and validated bioanalytical methods. Bioanalytical methods and techniques are continuously being changed and improved. It is also important to stress that each bio-analytical technique's specific characteristics differ from analysis to analysis. For each study, particular requirements for validation can be required. When testing samples for a given study at more than one site, the bioanalytical methods must be validated at each site and validated in the various areas for interlaboratory reliability must be adequately provided.[73].

Linearity

Linearity evaluates the system's ability to produce test results that are directly proportional to the sample analysis. Regardless of the process of drug production, the linear range of the procedure must be calculated. During the accuracy analysis, the starting and ending levels must be based on the five concentration levels. [74]. The following concentration levels are recommended for evaluation during method validation by ICH guidelines:

Assay: 80% –120% of concentrations of samples [finished product or pharmaceutical substances]. However, this range needs to be based on precise analysis. The linearity should be increased to a minimum of 75-125 percent of the nominal value, and accuracy should be prepared at 80, 100, and 120% of the nominal value.

Contents uniformity: 70 to 130% of the sample concentration is based on the type of dosage unless a more comprehensive, more adequate range is justified; [e.g., metered dose inhalers].

Dissolution method: 20% of the specified range is needed. If dissolution profiles are expected, the linearity range should begin at less than 120 percent of the total drug content recovered during the initial stage.

Impurity detection method: the degree of reporting is 120 percent.

Impurity and testing are combined in this form: a standard of 100 percent is used to quantify and reveal impurity at 120 percent of the test specification.

Accuracy

The accuracy of an analysis method is known as the degree of agreement between the value taken as a typical true or agreed on reference value and the value found. [75 76]

Bias

Bias may be expressed as a percentage deviation from an agreed-upon reference value. The term "trueness" refers to the mean value of a generally accepted reference value for a variety of measurements. It can be described in terms of partiality. Due to the high workload associated with evaluating such a complete sequence, truthfulness is usually not determined during process validation but rather from the results of numerous QCS during routine application [77].

precision

Precision is a term that refers to the degree to which a set of measurements are compiled under specific conditions from several samples of the same homogeneous sample. Precision is further subdivided into interday, intraday, and different analysts. Precision or repeatability tests have been carried out to assess accuracy in time. The use of different observers, instruments, reagents, and laboratories may have included these controls.[78].

Intermediate precision

In laboratories, the term "intermediate precision" applies to variations: different days, observers, and equipment, for example. [79] The ISO description referred to the expression "intermediate precision m-factor varies" when the m-factor denotes the number of variables that change between determinations [operator, equipment, or time]. Occasionally, intermediate precision is referred to as intermediate precision [80] between, between, or between days.

limit of detection [LOD]

The LOD is fully dependent on the boundary test. The smallest amount of analysis in a sample can be detected but not necessarily quantified under the given experimental conditions. The term "detection" is often used in conjunction with percentages, components per million, or parts per billion.

Limit OF Quantification [LLOQ]

LLOQ is a small sample quantity of study which can be calculated with appropriate precise and accurate measurements. LLOQ based on accuracy can be chosen as the most practical approach. The LLOQ is the lowest sample concentration but can be reliably and accurately quantified. For example,

chromatographical methods are used LLOQ only when basic noise is used according to signal and noise ratios. [81]

Robustness

According to the ICH guidelines, a robust analytical procedure is measuring its capacity to maintain its reliability during regular use without being influenced by minor but deliberate differences in process parameters [82-84]. The ability to replicate the [analytical] technique in various lab sites or other conditions, and the robustness test, which is an experimental set-up to determine the robustness of a system, without the incidence of uneven variations between the results achieved, can be identified.

Ruggedness

This covers various analysts, labs, columns, instruments, reagent sources, chemicals, and solvents. The research process's ruggedness is the degree of reproductiveness of test results obtained under many standard test conditions by analyzing the same samples [85]

Recovery study

Although the recovery study cannot be perfect, the scope of the analytes recovered and the internal norm should be consistent, reliable, and reproducible. For the extracted samples' analytics at three concentrations, recovery tests should be performed using unextracted parameters that indicate 100 percent of reconstruction [low, medium, and high]. [86-88]

Matrix effect

The matrix effect is defined as the effect of the co-eluting residual matrix of a biological sample on the ionization of the target. The matrix effect may be caused by organic and inorganic substances such as amines, urea, and carbohydrates. The variance in the coefficients calculated from a six-lot matrix is less than 15% of the normalized matrix factor [mf]. This can be accomplished at both low and high concentrations [up to three times the lower LLOQ] and near to the upper quantification limit [ULOQ]. The concentration is calculated to have an average coefficient of variation [CV] of less than 15%. [89-91]

TABLE-7 VALIDATION OF PARAMETERS AS PER ICH [GUIDELINES Q2A

S.NO	Parameter	Standard values
1	Accuracy	Recovery 98-102% with

		80,100,120% spiked sample.
2	Precision	RSD < 2%
2a	Repeatability	
2b	Intermediate precision	
3	Specificity/ selectivity	Interference <0.5%
4	Detection Limit	S/N > 2 or 3
5	Quantitation Limit	S/N > 10 ,RSD<20%
6	Linearity	r > 0.999
7	Range	80-120%
8	Stability	>24h or > 12h
9	Matrix effect	Matrix effect less than 100 indicates suppression matrix effect larger than 100is assign of matrix enhancement

Stability

The stability of the analyte is a necessary precondition for accurate quantification in the analysis process. The complete validation of a system must also involve stability tests at the various research steps, including pre-analysis storage. [92]

Sustainable.

Thawstability

In-processstability

SPECIFIC RECOMMENDATION FOR BIOANALYTICAL METHOD VALIDATION

A simple model is used to accurately explain the concentration/response relationship using appropriate weighting and fitness statistical tests. At least five concentration-level determinations can determine precision and accuracy for the validity of the bioanalytical process [excluding blank samples]. Except for LLOQ, the meaning is not more than 20 percent different from the theoretical meaning. The precision of around the medium value, except for the LLOQ not exceeding 20 percent of the CV, shall be no more than 15 percent of the CV. It may also be necessary to use other methods of assessing precision and accuracy that meet these constraints. [93] Re-injection reproducibility should be established to decide if an analysis interrupted by a fault of the device could be restarted

Documentation for method establishment

1.The analytical method's definition should be included in the method's implementation and establishment documents.

2.validation of pure drug criteria and their identity, as well as metabolite and internal standards used in validation studies[94].

3.Experiments were conducted to ensure that they were reliable, valid, recoverable, selective, quantity limit, and calibration curve, and that appropriate data was collected from these tests. [Equations and, if applicable, weighting functions]

4.Accuracy and precision of intra-and inter-testing are recorded.

5.Where applicable, details about cross-validation data in NDA [new drug approval] submissions.

6.Any deviations from the GLPS [as applicable] and justifications for such deviations, standard operating procedures, protocols, or [good laboratory practice]

Other information

The following may be included: abbreviations and other codes used such as study requirements, integration and reporting codes, reference lists, and copies of all sources that are readable. In these areas, standard operating procedures or protocols are provided:

1.Uniform calibration of acceptance or rejection requirements

2.Calibration of the acceptance or rejection conditions curves

3.Acceptance or rejection requirements for QC samples and assay runs

4.Acceptance criteria for observed values during the double-checking of all unknown samples

5.Coding classification, including clinical and preclinical sample codes, as well as sample codes for bioassays

6.Testing of clinical or preclinical batches

7.Sampling, processing, and storage of samples and
8.Repetitive sample studies.

APPLICATION OF THE VALIDATED METHOD TO ROUTINE DRUG ANALYSIS [95-96]

In the time available for the collection of stability information, the two samples in the biological matrix should be evaluated. In the case of biological samples, a single study without duplication or replicating analysis will generally be used when adequate variability is shown by the validation results in the test process. This declaration applies

to procedures that consistently comply with appropriate accuracy and precision tolerances.

Response function The standard curve should typically use the same fitting, weighting, and fitness defined during pre-study validation. Proper statistical testing based on the actual standard points shall decide the response feature during each validation. There are possible issues with the changes in response functions' connection from pre-study validation to routine running validation. To accept or deny the sprint, QC samples should be used. The matrix of these QC samples is spiked by the analyte.

System suitability: a particular standard operating procedure should be selected based on the analysis and methodology to ensure optimal functioning of the system.

Repeat analysis: For repeat and acceptance requirements, it is necessary to have a standard operating procedure in place. In this normal protocol or guideline, the rationale for repeating the sample analysis should be clarified. Repeated testing can involve repeated examination of clinical or pre-clinical samples for regulation, incoherence in duplicate testing, non-testing samples, sample processing errors, failure of equipment, weak chromatogram, and inconsistent pk efficiency. If the sample volume permits, reassays should be carried out three times. It is necessary to understand the reasoning for repeated study and the reporting of repeat analysis

Reintegration of sample data: a sop or a guideline should be provided for the sample data reintegration. The reasons for reintegration are how to carry out reintegration should be outlined in this sop or guideline. It is necessary to clearly explain and record the reason for reintegration. Reports should be made of actual and reintegration results.

CONCLUSION

I conclude that bioanalytical methods are commonly used to quantify drugs and metabolites in physiological matrices and that the methods may apply to studies in human clinical pharmacology and nonhuman pharmacology/toxicology. The bioanalytical method used to quantify drugs and their metabolites in biological fluids is important in the evaluation and interpretation of bioequivalence, pharmacokinetic [PK], and toxic kinetic studies. Low detection limits, the ability to produce structural information, the need for minimal sample

preparation, and the ability to cover a wide range of analytes with varying polarities are all advantages of LCMS-MS. Despite their high sensitivity and selectivity, LC/MS/MS instruments have some limitations due to matrix-induced variations in ionization efficiencies and ion suppression/enhancement effects caused by biological matrix.

Bibliography

1. Mohammad Mahdi Moeina, Aziza El Beqqali b, Mohamed Abdel-Rehim. Bioanalytical method development and validation: Critical concepts and strategies, Journal of Chromatography B, 1043 (2017) 3–11
2. D. Zimmer, Introduction to quantitative liquid chromatography-tandem mass spectrometry (LC-MS-MS), Chromatographia 57 (2003)325–332.
3. J. Schuhmacher, D. Zimmer, F. Tesche, V. Pickard, Matrix effects during an analysis of plasma samples by electrospray and atmospheric pressure chemical ionization mass spectrometry: practical approaches to their elimination, Rapid Commun. Mass Spectrom. 17 (2003)1950–1957.
4. Singh, A., Srinivasan, A.K., Chakrapani, L.N. and Kalaiselvi, P., 2019. LOX-1, the common therapeutic target in hypercholesterolemia: a new perspective of antiatherosclerotic action of aegeline. Oxidative medicine and cellular longevity, 2019.
5. T.M. Annesley, Ion suppression in mass spectrometry, Clin. Chem. 7 (2003) 1041–1047.
6. B.K. Matuszewski, M.L. Constanzer, C.M. Chavez-Eng, Strategies for the Assessment of Matrix Effect in Quantitative Bioanalytical Methods Based on HPLC–MS/MS, Anal. Chem. 75 (2003) 3019– 3030.
7. Singh, A., Gowtham, S., Chakrapani, L.N., Ashokkumar, S., Kumar, S.K., Prema, V., Bhavani, R.D., Mohan, T. and Sathyamoorthy, Y.K., 2018. Aegeline vs Statin in the treatment of Hypercholesterolemia: A comprehensive study in rat model of liver steatosis. Functional Foods in Health and Disease, 8(1), pp.1-16.
8. A. V. Eeckhaut, K. Lanckmans, S. Sarre, I. Smolders, Y. Michotte, Validation of bioanalytical LC-MS/MS assays: Evaluation of matrix effects, J. Chromatogr. B, 877 (2009)2198–2207.
9. M. Ahnoff, A-C. Nyström, F. Schweikart, A. Ekdahl, Matrix effect explained by the unexpected formation of peptide in acidified plasma, Bioanalysis 7 (2015)295-306
10. M.W.J. van Hout, H.A.G. Niederländer, R.A. de Zeeuw, G.J. de Jong, Ion suppression in the determination of clenbuterol in urine by solid-phase extraction atmospheric pressure chemical ionization ion-trap mass spectrometry, Rap. Com. Mass Spec. 17 (2003) 245–250.
11. D. Remane, M.R. Meyer, F.T. Peters, D.K. Wissenbach, H.H. Maurer, Fast and simple procedure for liquid-liquid extraction of 136 analytes from different drug classes for the development of aliquidchromatographic tandem mass spectrometric quantification method in human blood plasma, Anal. Bioanal. Chem. 397(2010) 2303–2314
12. Rahul Singh,¹ Ashok K. Shaky,² Rajashri Naik,² and NaeemShalan stability-indicating HPLC Determination of Gemcitabine in Pharmaceutical Formulations Hindawi Publishing Corporation International Journal of Analytical Chemistry Volume 2015, Article ID 862592, 12pages
13. Singh, A., Kumar, A. and Kalaiselvi, P., 2018. Aegeline, targets LOX1, the receptor for oxidized LDL to mitigate hypercholesterolemia: a new perspective in its anti-atherosclerotic action. Free Radical Biology and Medicine, 128, p.S41.
14. ShobhanaK.Menon*,BhoomikaR.Mistry,K uldeepV.Joshi,PinkeshG.Sutariya,Ravindra V. Patel Analytical detection and method development of anticancer drug Gemcitabine HCl using gold nanoparticles SpectrochimicaActa Part A: Molecular and

- Biomolecular Spectroscopy
15. Syed. Afrin, P. Prachet, Shaik. Munawar, Rama Rao. Nadendla Analytical Method Development and Validation of Gemcitabine in Tablets by HPLC by Different Analytical Techniques IOSR Journal Of Pharmacy And Biological Sciences (IOSR-JPBS) e-ISSN:2278-3008, p-ISSN:2319-7676. Volume 15, Issue 1 Ser. I (Jan –Feb 2020), PP24-29
 16. Pawanpreet Kaur^{1*} and Baljeet Singh analytical method development and validation of Gemcitabine: a review Indo American journal of pharmaceutical sciences coden [USA]: IAJPPB ISSN: 2349-7750
 17. Singh, A., 2022. Role of microbial metabolites in cardiovascular and human health. In *Microbiome, Immunity, Digestive Health and Nutrition* (pp. 137-148). Academic Press.
 18. Konatham Teja Kumar Reddy, & M. Akiful Haque. (2022). Develop and validate a highly sensitive method for the estimation of Molnupiravir in rat plasma by high-performance liquid chromatography-tandem mass spectrometry and its application to pharmacokinetic studies. *Journal of Pharmaceutical Negative Results*, 28–34. <https://doi.org/10.47750/pnr.2022.13.S01.0>
 19. Konatham Teja Kumar Reddy, Penke Vijaya Babu, Rajinikanth Sagapola, & Peta Sudhakar. (2022). A REVIEW OF ARTIFICIAL INTELLIGENCE IN TREATMENT OF COVID-19. *Journal of Pharmaceutical Negative Results*, 254–264. <https://doi.org/10.47750/pnr.2022.13.S01.31>
 20. Konatham Teja Kumar Reddy, Kumaraswamy Gandla, Penke Vijaya Babu, M Vinay Kumar Chakravarthy, Pavuluri Chandrasekhar, & Rajinikanth Sagapola. (2022). A CRITICAL REVIEW ON BIOANALYTICAL METHOD DEVELOPMENT AND VALIDATION OF FEW ONCOLOGY DRUGS BY USING LC-MS-MS. *Journal of Pharmaceutical Negative Results*, 16–27. <https://doi.org/10.47750/pnr.2022.13.S01.03>
 21. Reddy, K. T. K., & Haque, M. A. (2022). Bioanalytical method development and validation of atrasentan in human plasma using verapamil as internal standard by liquid chromatography coupled with tandem mass spectrometry. *International Journal of Health Sciences*, 6(S8), 625–638. <https://doi.org/10.53730/ijhs.v6nS8.10470>
 22. Konatham Teja Kumar Reddy et.al High Performance Liquid Chromatography for The Simultaneous Estimation of Anti-Ulcer Drugs in Pharmaceutical Dosage Form, *Journal of Positive School Psychology*, Vol. 6, No. 9, 4524-452
 23. Reddy KTK, Haque MA. Development and Validation of a High Throughput LC-MS/MS Method for Quantitation of Ipilimumab in Human Plasma. *International Journal of Pharmaceutical Quality Assurance*. 2022;13(3):303-307
 24. Teja Kumar Reddy Konatham, M. Anuradha (2020), a stability indicating method development and validation of Telmisartan and Nifedipine in pure form using RP-HPLC. *International Journal of Pharmaceutical, Biological and Chemical Sciences*, 9(3): 36-44
 25. Teja Kumar Reddy Konatham, Satyanarayana Reddy K., Anuradha Manipogo, a Review on viruses that originated from china; Sars, mers and covid-19 *World Journal of Pharmaceutical Research*, Vol 9, Issue 5, 2020, 2010-2015.
 26. Teja Kumar Reddy Konatham et al, A Systematic Review on Method Development and Validation of Few Antiviral Drugs by Using RP-HPLC. *Ijppr.Human*, 2021; Vol. 21 (3): 651-661.
 27. Konatham Teja Kumar Reddy and Kumaraswamy Gandla. Novel Vesicular Drug Delivery Systems Proniosomes. *Pharm Res* 2022, 6(3): 000272.
 28. Roh, J., Hill, J.A., Singh, A., Valero-Muñoz, M. and Sam, F., 2022. Heart failure with preserved ejection fraction: heterogeneous syndrome, diverse preclinical models. *Circulation Research*, 130(12), pp.1906-1925.

29. Asha Deepti et al / Stability Indicating Rp-Hplc Method For Assay Of Dexamethasone In Its Formulation, NEUROQUANTOLOGY, Stability Indicating Rp-Hplc Method For Assay Of Dexamethasone In Its Formulation| SEPTEMBER 2022 | VOLUME 20 | ISSUE 11 | PAGE 80-86| DOI: 10.14704/NQ.2022.20.11.NQ66009
30. Deepti, A., & Sahithi, A. (2020) Sensitive bioanalytical method development and validation of afatinib in human plasma by LC-ESI-MS/MS. *International Journal of Health Sciences*, 6(S5), 1736–1784. <https://doi.org/10.53730/ijhs.v6nS5.9522>
31. Kumar, K. P., Marathakam, A., Patnaik, S., Kumar, S., Sahithi, A., Priya, D. K. S., & Dogra, P. (2022). An insight of development and validation of bioanalytical method in the reference of anticancer drugs by using LC-MS/MS. *International Journal of Health Sciences*, 6(S3), 6349–6361. <https://doi.org/10.53730/ijhs.v6nS3.7454>
32. Sahithi et al., Stability indicating reverse-phase high-performance liquid chromatography method development and validation of imatinib, *International Journal of Research and Analytical Reviews*, Vol 7, Issue 3, 322-333, September 2020. DOI: 10.20959/wjpr20178-9069
33. Sahithi et al., Alapati Sahithi, B.Parijatha, D.Santhoshi Priya, Krishna Mohan Chinnala, Method Development and validation of Metformin by using RP-HPLC, *World Journal of Pharmaceutical Research*, 2277-7105, 6(8); August, 2017. DOI: 10.20959/wjpr20178-9069
34. K. Prashanthi*, E.Mounika, A. Sahithi, A. Chaitanya, C. Krishna Mohan Method Development and Validation for Determination of Dihydroergotamine in Dihydroergotamine Mesylate Nasal Spray Bidose by RP-HPLC, *Journal of Xidian University (Scopus)*, 1001-2004, 16(4) Apr 22. [Doi.org/10.37896/jxu16.4/042](https://doi.org/10.37896/jxu16.4/042) ISSN No:1001-2400
35. 30.separations7030047 Cayman chemicals product information Ruxolitinib item number11609
36. Anjana Vaghela, Ashok Patel, Ajay Patel, Amit Vyas, Nilesh Patel Sample Preparation In Bioanalysis: A review international journal of scientific & technology research volume 5, issue 05, May 2016 ISSN2277-8616
37. Wells DA. Fundamental strategies for bioanalytical sample preparation. High throughput bioanalytical sample preparation methods and automation strategies. Amsterdam: Elsevier; 2002. p. 41
38. Xu X. Book Using mass spectrometry for drug metabolism studies. In: Korfmacher W, editor. Chapter 7: Fast Metabolite Screening in a Discovery Setting. 2nd ed. CRC Press. 2010
39. Churchwell MI, Twaddle NC, Meeker LR, Doerge DR. Improving LCMS sensitivity through increases in chromatographic performance: Comparison of UPLC-ES/MS/MS to HPLC-ES/MS/MS. *J Chromatogr B Analyt Technol Biomed Life Sci* 2005;825:134-43.
40. sample preparation for bioanalysis 2012 evaluation group co and R @2011 water corporation
41. John Ayrton, ph.d., GlaxoSmithKline quantitative drug analysis and pharmacokinetics make synergistic partnership water the science of what's possible
42. Wenkui Li, Wenying Jian, and Yunlin Fu Protein Precipitation, Liquid-Liquid Extraction, and Solid-Phase Extraction Basic Sample Preparation Techniques in LC-MS Bioanalysis
43. Sarika Khasnis. Large Molecule Bioanalysis Outsourcing opportunities & Challenges. *Pharma Focus Asia Magazine*
44. Shiv Chandra Singh, A., Yu, A., Chang, B., Li, H., Rosenzweig, A. and Roh, J.D., 2021. Exercise Training Attenuates Activin Type II Receptor Signaling in the Aged Heart. *Circulation*, 144(Suppl_1), pp.A14259-A14259.
45. Bulletin 910 Guide to Solid Phase

- Extraction.<http://www.supelco@sial.com/>
©1998 Sigam- Aldrich Co. Page No.-3
[collected2011.10.10]
46. P. Wal, B. Kumar, Dr.A. Bhandari, A.K. Rai, Ankit Wal. Bioanalytical Method Development- Determination of Drug in Biological Fluids. Journal of Pharmaceutical Science and technology vol.2 (10)210,333-347
 47. P.LaxmanKole, G. Venkatesh, J. Kotechacand R. Sheshalad. Recent advances are β sample preparation techniques for effective bioanalytical methods. Biomedical Chromatography. 2011;25
 48. K.Möller. Molecularly Imprinted Solid-Phase Extraction and Liquid Chromatography/Mass Spectrometry for Biological Samples Doctoral Thesis, Department of Analytical Chemistry Stockholm University2006
 49. L.R. Snyder, JJ Kirk. Sample preparation; Practical HPLC method development, 2nd edition, John Wiley and Sons Inc.; 2000:110-141
 50. method of chemical analysis 5th edition Himalaya publication. G. R. Chatwal, s.k. Anand. Instrumental http://en.wikipedia.org/wiki/pharmacokinetic_studies [collected2011.12.1]
 51. XiaobinGongat, Le Yangat, Feng Zhanga, YoutianLianga, ShouhongGaoa, KeLiub*, Wansheng Chen, Validated UHPLC-MS/MS assay for the quantitative determination of etoposide, gemcitabine, vinorelbine and their metabolites in lung cancer patients
 52. Tashinga E. Bapiro · Frances M. Richards · Mae A. Goldgraben · Kenneth P. Olive · Bassetti Madhu · Kristopher K. Frese · Natalie Cook · Michael A. Jacobetz · Donna-Michelle Smith · David
 53. A. Tuveson · John R. GriYths · Duncan I. Jodrell A novel method for quantification of gemcitabine and its metabolites 2,2-diXuoodeoxyuridine and gemcitabine triphosphate in tumor tissue by LC-MS/MS: comparison with ^{19}F NMR Cancer ChemotherPharmacol (2011) 68:1243–1253 DOI 10.1007/s00280-011-1613-0spectroscopy
 54. Ling-Zhi Wang,1,2 Wei-Peng Yong,1 Ross-A. Soo,1 Soo-Chin Lee1, Richie Soong2, HowSung Lee,3 Boon-Cher Goh, Rapid Determination of Gemcitabine and Its Metabolite in Human Plasma by LC-MSMS through Micro Protein Precipitation with Minimum Matrix Effect Ling-Zhi Wang et al /J. Pharm. Sci. & Res. Vol.1(3), 2009, 23-32ISSN:0975-1459
 55. DP, S. K., P. C, and S. R. Jvln. "development and validation of a dried blood spot lc-ms/ms assay to quantify gemcitabine in human whole blood: a comparison with and without cytidine deaminase inhibitor". InternationalJournal of Pharmacy and Pharmaceutical Sciences, Vol. 8, no. 8, Aug. 2015, pp. 75-81,
 56. S. A. Veltkamp,1,2* M. J. X. Hillebrand,3 H. Rosing,3 R. S. Jansen,3 E. R. Wickremsinhe,5 E. J. Perkins,5 J. H. M. Schellens1,2,4 and J. H. Beijnen2,3,4Quantitative analysis of Gemcitabine triphosphate in human peripheral blood mononuclear cells using weak anion-exchange liquid chromatography coupled with tandem mass spectrometry journal OF MASS SPECTROMETRY J. Mass Spectrom. 2006; 41: 1633–1642 Published online 20 November 2006 in Wiley InterScience (www.interscience.wiley.com) DOI: 10.1002/jms.1133
 57. Michael T. Furlong,*, Shruti Agrawal b, Dara Hawthornea, Michael Lagoc, Steve Unger d, Linda Krueger d, Bruce Stouffer A validated LC-MS/MS assay for the simultaneous determination of the anti-leukemic agent dasatinib and two pharmacologically active metabolites in human plasma: Application to a clinical pharmacokinetic study journal of Pharmaceutical and Biomedical Analysis 58 (2012) 130–135
 58. Hadir M. Maher1,2*, Nourah Z. Alzoman1, Shereen M. Shehata1, Norah O. Abanmy Validated UPLC-MS/MS method for the quantification of dasatinib in plasma: Application to pharmacokinetic interaction studies with nutraceuticals in Wistar rats

- PLoS ONE |
<https://doi.org/10.1371/journal.pone.0199208>
 08 June 14, 2014
59. Prinesh N. Patel, a
 GananadhamuSamanthula,*a
 VeeraraghavanSridhar,bRambabuArla,bK
 anthi Kiran V. S. Varanasib and Swaroop
 Kumar V.V.SbValidated LC-MS/MS
 method for simultaneous determination of
 Dasatinib and Sitagliptin in rat plasma and
 its application to pharmacokinetic study
 This journal is © The Royal Society of
 Chemistry 2014 Anal. Methods, 2014, 6,
 433–439
 60. S. R. S. Thappali 1, K. V. S. Varanasi 1, S.
 Veeraraghavan 1, S. K. V. S. Vakkalanka 1,
 M. Khagga 2Simultaneous Determination
 of Methotrexate, Dasatinib, and its Active
 Metabolite N- Deshydroxyethyl Dasatinib
 in Rat Plasma by LC-MS/ MS: Method
 Validation and Application to
 Pharmacokinetic Study, Thappali SRS et
 al. Simultaneous determination of
 methotrexate... Arzneimittelforschung
 2012; 62:624–630
 61. Wen1, q. Zhang2, y. He1, m. Deng1, x.
 Wang1, and j. Ma3,* gradient elution lc-ms
 determination of Dasatinib in Rat Plasma
 and Its Pharmacokinetic Study 0231–2522
 © 2013 Akadémiai Kiadó, Budapest
 62. Dai G, Pfister M, Blackwood-Chirchir A,
 Roy A. Importance of characterizing
 determinants of variability in exposure:
 application to dasatinib in subjects with
 chronic myeloid leukemia. J
 Clin Pharmacol 2008;48:1254e69
 63. De Francia S, D'Avolio A, De Martino F,
 Pirro E, Baietto L, Siccardi M, et al. New
 HPLC/MS method for the simultaneous
 quantification of the antileukemia drugs
 imatinib, dasatinib, and nilotinib in human
 plasma. J Chromatogr B Anal Technol
 Biomed Life Sci 2009;877:1721e6.
 64. Haouala A, Zanolari B, Rochat B,
 Montemurro M, Zaman K, Duchosal MA,
 et al. Therapeutic drug monitoring of the
 new targeted anticancer agents imatinib,
 nilotinib, dasatinib, sunitinib, sorafenib,
 and lapatinib by LC tandem mass
 spectrometry. J Chromatogr B Anal
 Technol Biomed Life Sci 2009;877:1982e96
 65. Bouchet S, Chauzit E, Ducint D, Castaing
 N, Canal-Raffin M, Moore N, et al.
 Simultaneous determination of nine
 tyrosine kinase inhibitors by 96-well solid-
 phase extraction and ultra-performance
 LC/MS/MS. Clin Chim Acta 2011;412:1060e7
 66. Boini, K.M., Singh, A. and Koka, S.S.,
 2021. Gut Microbial Metabolite
 Trimethylamine N-oxide Enhances
 Endoplasmic Reticular Stress and
 Promotes Endothelial Dysfunction.
 Circulation, 144(Suppl_1), pp.A14071-
 A14071.
 67. Sridhar Veeraraghavana,c,*,
 SatheeshmanikandanThappali a,
 SrikantViswanadhaa,
 SandhyaraniChennupati a,
 SanthoshkumarNallaa,
 ManikantakumarGollaa,
 SwaroopkumarVakkalankaa,
 ManivannanRangasamySimultaneous
 quantification of ruxolitinib and nilotinib
 in rat plasma by LC-MS/MS: Application
 to a pharmacokinetic study journal of
 Pharmaceutical and Biomedical Analysis
 94 (2014) 125–131
 68. clairePressiat, PharmD, PhD,*† Huu-Hien
 Huynh, PharmD,* Alain Plé,*† Héléne
 Sauvageon, PharmD,*† Isabelle Madelaine,
 PharmD, PhD,† Cécile Chougnat, MD,‡
 Christine Le Maignan, MD,§
 Samia Mourah, PharmD, PhD,*¶ and
 Lauriane Goldwirt, PharmD,
 PhDDevelopment and Validation of a
 Simultaneous Quantification Method of
 Ruxolitinib, Vismodegib, Olaparib, and
 Pazopanib in Human Plasma Using Liquid
 Chromatography Coupled With Tandem
 Mass Spectrometry
 69. Vander HY, Nij huis A, Verbeke JS,
 Vandeginste BG, Massart DL. Guidance
 for robustness/ruggedness test in method
 validation. J Pharm Biomed
 Anal 2009;24:723-53.
 70. Kringle RO. An assessment of the 4-6-20
 rule of acceptance of analytical runs in
 bioavailability, bioequivalence, and
 pharmacokinetic studies. Pharm

- Res1994;11(4):556-60.
71. Wieling J, Hendriks G, Tamminga WJ, Hempenius J, Mensink CK, Oosterhuis B, et al. Rational experimental design for bioanalytical methods validation. Illustration using an assay method for total captopril in plasma. *J Chromatogr A*1996;730(1-2):381-94.
72. Kringle R, Hoffman D. Stability methods for assessing the stability of compounds in whole blood for clinical bioanalysis. *Drug Inf J*2001;35:1261-
73. Viswanathan CT, Bansal S, Booth B, DeStefano AJ, Rose MJ, Sailstad J, et al. Quantitative bioanalytical methods validation and implementation: Best practices for chromatographic and ligand binding assays. *AAPS J* 2007;9(2):E260-7
74. Mark H. Application of improved procedure for testing linearity of an analytical method to pharmaceutical analysis. *J Pharm Biomed Anal*2003;33(1):7-20
75. Puluido A, Ruusanches I, Boque R, Rius FX. Uncertainty of results in routine qualitative analysis in analytical chemistry. *J Pharm Biomed Anal*2005;22:647-54
76. Singh PS, Shah G. Analytical method development and validation. *J Pharm Res*2011;4(5):2330-2
77. Singh UK, Pandey S, Pandey P, Keshri PK, et al. (2008) Bioanalytical method development and validation. *ExpressPharma*.
78. Murugan S, Pravallika N, Sirisha P, Chandrakala K (2013) A Review on Bioanalytical Method Development and Validation by Using LC-MS/MS. *J of Chem and Pharma Sci* 6:41-45.
79. McPolin O (2009) Validation of an analytical method for the pharmaceutical analysis, Mourne trainingservices
80. Sekar V, Jayaseelan S, Subash N, Kumar EU, Perumal P, et al. (2009) Bioanalytical Method Development and Validation of Letrozole by Rp-HPLC Method. *Int J of Pharm Res and Develop* 1: 1- 8.
81. Vinod P. Shah, Bioanalytical Method Validation, A Revisit with a Decade of Progress, *Pharmaceutical Research*, December 2000, Volume 17, Issue 12, pp 1551–155722.
82. David Watson G., *Pharmaceutical Analysis* (3 rd Ed., Churchill Livingstone, London: Harcourt Publishers Limited, Essex CM 20 2JE,2012
83. Current Good Manufacturing Practices for Finished Pharmaceuticals, 21 CFR, Parts 210 and 211, US Food and DrugAdministration.
84. Willard HH, Merritt Jr LL, Dean JA, Settle Jr FA. *Instrumental methods of analysis*.CBS Publ.; 1981:19-90
85. McDowall RD. The role of laboratory information management systems LIMS in analytical method validation. *Anal ChimActa* 2007; 54:149-5.
86. Wieling J, Hendriks G, Tamminga WJ, Hempenius J, Mensink CK, Oosterhuis B, et al. Rational experimental design for bioanalytical methods validation. Illustration using an assay method for total captopril in plasma. *J Chromatogr A* 1996;730(12):381-9
87. Bmscheck T, Meyer H, Wellhrner HH. A High-performance liquid chromatographic assay for the measurement of azathioprine in human serum samples. *J Chromatogr*1996;212:287-94.
88. Kees F, Jehnich D, Grobecker H. Simultaneous determination of acetylsalicylic acid and salicylic acid in human plasma by high-performance liquid chromatography. *J Chromatogram*1996;677:172-7.
89. Raymond NX, Fan LR, Matt hew J, Tawakol A. Recent advances in high-throughput quantitative bioanalysis by LC-MS/MS. *J Pharm Biomed Anal*2007;44:342-55.
90. Lau Y, Hanson GD, Carel BJ. Determination of rifampin in human plasma by HPLC with ultraviolet detection. *J Chromatogr*1998;676:147-52
91. Hartmann C, Massart D, McDowall RD. An analysis of the Washington Conference Report on bioanalytical method validation. *J Pharm Biomed Anal* 2005;12:1337-43.
92. Karnes HT, Shiu G, Shah VP. Validation of

- bioanalytical methods. Pharm Res 2001;8:421-6.
93. Lindner W, Wainer IW. Requirements for initial assay validation and publication in J ChromatographyB. J Chromatogr2006;707:1-2.
 94. Penninckx W, Hartmann C, Massart DL, Smeyers-Verbeke J. Validation of the Calibration Procedure in Atomic Absorption Spectrometric Methods. J Anal At Spectrom1998;11:237-46.
 95. Wieling J, Hendriks G, Tamminga WJ, Hempenius J, Mensink CK, Oosterhuis B, et al. Rational experimental design for bioanalytical methods validation. Illustration using an assay method for total captopril in plasma. J Chromatogr2006;730:381-94
 96. Dighe S, Shah VP, Midha KK, McGilveray IJ, Skelly JP, Yacobi A, et al. Analytical methods validation: bioavailability, bioequivalence, and pharmacokinetic studies. Conference Report. Eur J Drug Metabol Pharmacokinetics 1998;16:249-55.